

A New Niche for *Vibrio logei*, the Predominant Light Organ Symbiont of Squids in the Genus *Sepiola*

PAT M. FIDOPIASTIS,¹ SIGURD VON BOLETZKY,² AND EDWARD G. RUBY^{1*}

*Pacific Biomedical Research Center, University of Hawaii, Manoa, Honolulu, Hawaii,¹
and Laboratoire Arago, CNRS-URA2156-OOB, Banyuls-sur-Mer, France²*

Received 16 July 1997/Accepted 27 October 1997

Two genera of sepiolid squids—*Euprymna*, found primarily in shallow, coastal waters of Hawaii and the Western Pacific, and *Sepiola*, the deeper-, colder-water-dwelling Mediterranean and Atlantic squids—are known to recruit luminous bacteria into light organ symbioses. The light organ symbiont of *Euprymna* spp. is *Vibrio fischeri*, but until now, the light organ symbionts of *Sepiola* spp. have remained inadequately identified. We used a combination of molecular and physiological characteristics to reveal that the light organs of *Sepiola affinis* and *Sepiola robusta* contain a mixed population of *Vibrio logei* and *V. fischeri*, with *V. logei* comprising between 63 and 100% of the bacteria in the light organs that we analyzed. *V. logei* had not previously been known to exist in such symbioses. In addition, this is the first report of two different species of luminous bacteria co-occurring within a single light organ. The luminescence of these symbiotic *V. logei* strains, as well as that of other isolates of *V. logei* tested, is reduced when they are grown at temperatures above 20°C, partly due to a limitation in the synthesis of aliphatic aldehyde, a substrate of the luminescence reaction. In contrast, the luminescence of the *V. fischeri* symbionts is optimal above 24°C and is not enhanced by aldehyde addition. Also, *V. fischeri* strains were markedly more successful than *V. logei* at colonizing the light organs of juvenile *Euprymna scolopes*, especially at 26°C. These findings have important implications for our understanding of the ecological dynamics and evolution of cooperative, and perhaps pathogenic, associations of *Vibrio* spp. with their animal hosts.

Members of at least two families of squids and seven families of marine fishes are known to harbor a monospecific culture of luminous bacteria in specialized light organs (23, 29). In these cooperative symbioses, the host provides the bacteria with nutrients and a protected environment in exchange for light production, which the host may use for intraspecific communication, prey attraction, or predator avoidance (22). To date, only three species (*Vibrio fischeri*, *Photobacterium leiognathi*, and *Photobacterium phosphoreum*) of the 10 described marine luminous bacteria have been isolated from light organs. *P. leiognathi* and *P. phosphoreum* occur primarily as symbionts of marine fishes (14, 28). *V. fischeri* has been identified as the specific light organ symbiont of at least five species of marine animal hosts, including several bobtail squids in the family Sepiolidae (4, 29) and fishes in the family Monocentridae (10, 31). In all cases, the light organs of marine animals have been reported to contain only a single species of luminous bacteria (23).

Euprymna and *Sepiola* are two important genera in the family Sepiolidae (19). *Euprymna* spp. are found primarily in the shallow coastal waters of the Pacific Ocean, in Hawaii (*Euprymna scolopes*), Australia (*Euprymna tasmanica*), and the Indo-West Pacific region (*Euprymna morsei*). The light organ symbiont of these squids has been identified as *V. fischeri* (6, 18, 24, 30). In contrast, members of the genus *Sepiola* are found in the Mediterranean Sea, along the Atlantic coastline of Europe, and also in Japan and the Philippines. Until now, the identity of the light organ symbionts of these relatively deeper- and colder-water *Sepiola* spp. had been reported only for *Sepiola atlantica* (13).

Preliminary biochemical and microscopic analyses of light organ symbionts isolated from the Mediterranean squids *Sepiola affinis* and *Sepiola robusta* suggested that these bacteria were either *V. fischeri* or the closely related luminous species *Vibrio logei*. While these two bacteria have proven difficult to differentiate by conventional phenotypic traits (2, 23), a substantial genomic sequence difference does exist among *Vibrio* spp. in a variable region of the 16S rRNA gene called V1 (36). The length of a PCR-amplified 16S rRNA gene (rDNA) fragment containing this region can be used to categorize different members of the genus *Vibrio* (33).

We used this PCR product analysis, combined with 16S rDNA sequence information and other phenotypic data, to determine that the predominant light organ symbionts of *S. affinis* and *S. robusta* are *V. logei*. Surprisingly, we found that *V. fischeri* was a minor but significant component of the symbiont population in some of the light organs, suggesting that there may be a dynamic competition between these two luminous species for dominance. To test this hypothesis, we compared two ecologically relevant activities in representative isolates of *V. logei* and *V. fischeri* symbionts: (i) the effect of temperature on growth rate and luminescence potential, and (ii) their ability to colonize the light organs of juvenile *E. scolopes* squids.

MATERIALS AND METHODS

Collection of specimens of *Sepiola* spp. and isolation of light organ symbionts. Specimens of bobtail squids were collected from the upper 50 m of seawater within 1 km of shore near the Laboratoire Arago, Banyuls-sur-Mer, France, between 1 and 15 July 1995. The animals (mantle length, 13 to 24 mm) were identified as *S. affinis* and *S. robusta* by using the criteria outlined by Bello (4).

Bacterial symbionts were isolated from three specimens each of freshly collected *S. affinis* and *S. robusta* by dissecting an anesthetized animal, removing the light organ, and homogenizing it in sterile seawater. Aliquots of the homogenate were spread on seawater-tryptone-yeast extract (SWT) agar medium (6) and incubated at 25°C overnight. Uniform fields of golden-yellow colonies were observed on all plates, although some light organs yielded two distinct colony size

* Corresponding author. Mailing address: PBRC, University of Hawaii, 41 Ahui St., Honolulu, HI 96813. Phone: (808) 539-7309. Fax: (808) 599-4817. E-mail: eruby@hawaii.edu.

classes. Four to six representative colonies from each light organ (designated either SA for *S. affinis* symbionts or SR for *S. robusta* symbionts) were isolated and stored for further study.

PCR amplification of the V1 region of the 16S rRNA gene. Bacterial cultures of bobtail squid symbionts and reference strains (see Table 1) were streaked on SWT agar and incubated at 20°C overnight. Between 10^8 and 10^9 cells from single colonies were inoculated into tubes containing 1.5 ml of distilled water that had been filtered through 0.22- μ m-pore-size filters and autoclaved. The cells were lysed by exposure to five cycles of freezing in a dry ice–95% ethanol bath and thawing in a 37°C water bath. Cell debris was pelleted by centrifugation for 5 min. Each supernatant was transferred to a separate tube, and 1 μ l of this supernatant was added to 100 μ l of the PCR solution (containing 200 μ M each deoxynucleoside triphosphate, 10 mM Tris-HCl [pH 8.3], 50 mM KCl, 1.5 mM MgCl₂, 2.5 U of *Taq* polymerase, and 0.1 μ M each primer). Forward and reverse PCR primers used to specifically amplify V1 (33) were as follows: RU41 (forward) 5' GATCATGGCTCAGATTGAACG 3'; and RU42 (reverse), 5' AGG CATTACTCACCCGTCC 3'.

These PCR solutions were overlaid with sterile mineral oil and inserted into a thermal cycler for 37 cycles under the following conditions: 94°C, 6 min (first cycle only); 94°C, 1.5 min; 50°C, 1.5 min; 72°C, 2 min; and 72°C, 7 min (final cycle only). Aliquots of each PCR product were then loaded into separate wells of a 3% agarose gel and electrophoretically separated for 5 h at 60 V.

Cloning and sequencing of a portion of the 16S rRNA gene. Entire 16S rRNA genes from light organ symbionts of *S. affinis* (strains SA1, SA5, SA6, and SA12), and *S. robusta* (strains SR5 and SR6) were PCR amplified with the following primers: RU1 (reverse), 5' CCTTCCCTCACGGTACTGT 3'; and RU2 (forward), 5' TGGCTCAGATTGAACGCTGGCGG 3'.

PCR products from strains SA1, SA5, SA6, and SR5 were purified by passage through QIAquick columns (Qiagen Inc., Chatsworth, Calif.) and directly sequenced at the Biotechnology-Molecular Biology Instrumentation Facility, University of Hawaii, Manoa. Purified PCR products from strains SA12 and SR6 were ligated into the vector pCRII and cloned in *Escherichia coli* with the TA cloning kit (Invitrogen Corp., San Diego, Calif.) in accordance with the manufacturer's instructions. Plasmids containing the desired 2.3-kbp 16S rDNA inserts from strains SA12 and SR6 were then prepared for sequence analysis.

Denatured plasmid DNA (between 300 and 500 ng) was placed into an Eppendorf tube and mixed with 2 μ l of 5 \times sequencing buffer (0.2 M Tris-HCl, pH 7.5; 0.1 M MgCl₂; and 0.25 M NaCl) and primer RU2 (10 ng). The tubes were incubated at 37°C for 20 min followed by a 25°C incubation for 10 min to allow the primer to anneal to the 16S rDNA insert. Following this step, the 16S rDNA inserts were sequenced by using the Sequenase version 2.0 sequencing kit (U.S. Biochemicals, Cleveland, Ohio). Sequence data were analyzed with the BLAST algorithm (1) to identify sequences with a high degree of identity to our query sequences.

Biochemical tests for distinguishing *V. logei* from *V. salmonicida*. Because 16S rDNA sequence comparisons were not able to unambiguously differentiate between *V. logei* and *Vibrio salmonicida*, we subjected symbionts from *Sepiolo* spp. to three physiological tests that identify biochemical capabilities (lysine decarboxylation, D-galactose fermentation, and nitrate reduction) that differ for these two *Vibrio* species. The test media were prepared and the results were interpreted in accordance with the Difco Manual (7a), except that each medium was supplemented with marine cations according to the recipe of Farmer and Hickman-Brenner (9). Test organisms were pregrown on SWT agar at 14°C before inoculation, and the reaction tubes were incubated at the same temperature for between 48 and 96 h prior to the recording of results.

Effect of temperature on the growth rates of symbionts of *Sepiolo* spp. Bacterial strains were inoculated into flasks containing 20 ml of SWT broth to a final optical density (OD) at 600 nm of about 0.01 (approximately 5×10^7 cells/ml). The cultures were then shaken at either 20, 24, or 28°C, and their ODs were measured periodically throughout the logarithmic growth phase.

Effect of temperature and aldehyde on the luminescence of symbiont strains. Bacterial cultures were inoculated into flasks containing *Vibrio harveyi*-conditioned broth medium, prepared as described previously (20). This conditioning has been shown to remove an inhibitor of luminescence induction. The cultures were incubated with shaking at either 20, 24, or 26°C. During the logarithmic growth phase, 1.0-ml aliquots of each culture were removed at regular intervals, measured for luminescence, and then exposed to 5 μ l of a 0.00005% (vol/vol) decyl aldehyde suspension in water, and the level of luminescence was remeasured.

Colonization of the *E. scolopes* light organ by symbionts of *Sepiolo* spp. Bacterial strains to be tested for their ability to colonize juvenile *E. scolopes* were grown, without shaking, to an OD of between 0.1 and 0.3 in SWT broth at either 21 or 26°C, depending on the temperature at which the corresponding colonization experiment was to be performed. These cultures were diluted to about 2,000 cells per ml in natural seawater to serve as the inocula (17). Newly hatched juveniles of *E. scolopes* were placed into glass vials containing 5 ml of one of these seawater suspensions of bacteria. The juveniles were exposed to these inocula for 3 h and then washed and placed in 5 ml of natural seawater without added bacteria. The luminescence of the squids was subsequently monitored photometrically in the morning and at dusk for the next 2 to 4 days. After the final luminescence reading was taken, each animal was washed three times in sterile seawater, placed in an Eppendorf tube containing 700 μ l of sterile sea-

water, and homogenized with a pestle. Dilutions of the homogenate were spread in triplicate on SWT agar and incubated at 24°C for between 24 and 48 h. The resulting colonies were checked for luminescence, and estimates of the number of CFU per light organ were calculated.

RESULTS

Collection and identification of light organ symbionts of Mediterranean bobtail squids. All of the colonies arising from the highest dilutions of *Sepiolo* light organ homogenates had the golden-yellow color characteristic of *V. fischeri* and *V. logei*. From the number of CFU per ml of diluted homogenate, it was possible to calculate that each of the six light organs contained between 2×10^7 and 4×10^8 total symbionts. Interestingly, homogenates of two of the six bobtail squids (one specimen of *S. affinis* and one of *S. robusta*) gave rise to a mixture of two colony types, based on size and light emission. When observed in the dark, the larger class of colonies produced a relatively brighter bioluminescence. From the one *S. affinis* specimen, these larger colonies constituted about 37% of the total CFU, while in the *S. robusta* plating, they accounted for 19%. Five strains were isolated from these two less-abundant colony classes: SA1, SA2, SA3, SA4, and SR5 (Table 1). They and five representatives of the more abundant class, as well as 17 isolates from the other four light organs, were chosen for taxonomic identification.

Differentiation of *V. logei* and *V. fischeri* symbionts on the basis of V1 region size. The *V. logei* type strains ATCC 15382 and ATCC 35077, two luminous bacterial isolates from Southern California seawater (MdR7 and MdRD), and 22 of 27 of the isolates from *Sepiolo* spp. light organs each contained a 111-bp fragment when the portion of its 16S rDNA sequence that spanned the V1 region was PCR amplified (Fig. 1). In contrast, *V. fischeri* type strain ATCC 25918, the fish symbiont strain MJ1, four light organ isolates from *Euprymna* spp. (ES114, ES213, EM17, and EM24), and the remaining 5 of the 27 *Sepiolo* light organ symbionts each contained a 121-bp fragment (Fig. 1 and Table 1). These results support the use of this technique as a relatively easy method of distinguishing between *V. logei* and *V. fischeri* strains, and they provided the first indication that over 80% of the *Sepiolo* symbionts examined were most closely related to *V. logei* while the remainder were most closely related to *V. fischeri* (Table 1).

Confirmation of symbiont identification by partial 16S rDNA sequences and biochemical tests. To further confirm their identity and support the conclusions based on V1 size, partial 16S rDNA sequences from strains SA1, SA5, SA6, SA12, SR5, and SR6 were obtained. When analyzed against the GenBank nucleic acid database, the 16S rDNA sequences (between 250 and 800 bp in length) from SA1 and SR5 each aligned with the highest degrees of identity (90 and 92%, respectively) to *V. fischeri* 16S rDNA. Similarly, the 16S rDNA sequences of SA5, SA6, SA12, and SR6 aligned with the highest degrees of identity (93, 94, 97, and 99.6%, respectively) to *V. logei*. Thus, the V1 region size and partial 16S rDNA sequence comparisons were in agreement (Table 1).

During these analyses we discovered that the 16S rDNA sequence from *V. salmonicida* also aligned with a high (>90%) degree of identity to SA5, SA6, SA12, and SR6 and that *V. salmonicida* had a 111-bp V1 region. While the data suggested that these four symbionts were most closely related to *V. logei*, the relatively high degree of similarity to *V. salmonicida* produced some ambiguity. Thus, we examined the expression of several metabolic activities that differ between these two *Vibrio* species. Strains of *V. logei* and *V. salmonicida* were easily differentiated on the basis of the results of three biochemical tests

TABLE 1. Bacterial strains used in this study

Strain	Source (reference)	Size of V1 region ^a	Highest degree of 16S rDNA sequence similarity to:
ATCC 15382	Pacific cod skin (7)	S	<i>V. logei</i> ^b
ATCC 35077	Tanner crab lesion (3)	S	<i>V. logei</i> ^b
ATCC 25918	Seawater (26)	L	<i>V. fischeri</i> ^b
ES1114	<i>E. scolopes</i> light organ (6)	L	<i>V. fischeri</i> ^b
MJ1	<i>Monocentris japonica</i> light organ (31)	L	<i>V. fischeri</i> ^b
NCMB 2262	Atlantic salmon infection (8)	S	<i>V. salmonicida</i> ^b
MdR7	California coastal seawater (16)	S	<i>V. logei</i>
MdRD	California coastal seawater (16)	S	— ^c
SA1	<i>S. affinis</i> no. 1 (this study)	L	<i>V. fischeri</i>
SA2	<i>S. affinis</i> no. 1 (this study)	L	—
SA3	<i>S. affinis</i> no. 1 (this study)	L	—
SA4	<i>S. affinis</i> no. 1 (this study)	L	—
SA5	<i>S. affinis</i> no. 1 (this study)	S	<i>V. logei</i>
SA6	<i>S. affinis</i> no. 1 (this study)	S	<i>V. logei</i>
SA7	<i>S. affinis</i> no. 2 (this study)	S	—
SA8	<i>S. affinis</i> no. 2 (this study)	S	—
SA9	<i>S. affinis</i> no. 2 (this study)	S	—
SA10	<i>S. affinis</i> no. 2 (this study)	S	—
SA11	<i>S. affinis</i> no. 2 (this study)	S	—
SA12	<i>S. affinis</i> no. 2 (this study)	S	<i>V. logei</i>
SA13	<i>S. affinis</i> no. 3 (this study)	S	—
SA14	<i>S. affinis</i> no. 3 (this study)	S	—
SA15	<i>S. affinis</i> no. 3 (this study)	S	—
SA16	<i>S. affinis</i> no. 3 (this study)	S	—
SA17	<i>S. affinis</i> no. 3 (this study)	S	—
SA18	<i>S. affinis</i> no. 3 (this study)	S	—
SR1	<i>S. robusta</i> no. 1 (this study)	S	—
SR2	<i>S. robusta</i> no. 2 (this study)	S	—
SR3	<i>S. robusta</i> no. 2 (this study)	S	—
SR5	<i>S. robusta</i> no. 2 (this study)	L	<i>V. fischeri</i>
SR6	<i>S. robusta</i> no. 2 (this study)	S	<i>V. logei</i>
SR7	<i>S. robusta</i> no. 3 (this study)	S	—
SR8	<i>S. robusta</i> no. 3 (this study)	S	—
SR9	<i>S. robusta</i> no. 3 (this study)	S	—
SR10	<i>S. robusta</i> no. 3 (this study)	S	—

^a S, small (111-bp) V1 region; L, large (121-bp) V1 region.
^b Taxonomic criteria other than DNA sequence similarity were used to identify these strains (see references).
^c —, 16S rRNA gene sequencing was not performed for this strain.

(Table 2). These data indicate that although *V. salmonicida* and *V. logei* strains have very similar 16S rDNA sequences, they exhibit considerable metabolic divergence.

Differential effects of temperature on growth and luminescence of *V. logei* and *V. fischeri*. One of the few phenotypic traits that has been reported to be generally useful for distinguishing between *V. logei* and *V. fischeri* is the relative psychrophily of the former. Thus, it was not surprising that *V. logei* strains from either *S. affinis* or *S. robusta* grew well on SWT agar plates incubated for 4 days at 4°C whereas *V. fischeri* strains did not grow at all under those conditions (data not shown). Similarly, *V. logei* symbionts grew more rapidly in SWT medium at 20°C than their *V. fischeri* counterparts (Fig. 2A). In contrast, at temperatures above 20°C, the *V. fischeri* symbionts had a distinct growth advantage over *V. logei* symbionts (Fig. 2A). All *V. logei* SA and SR strains that were examined grew in culture with generation times between 1.3 and 2.2 times longer at 28°C

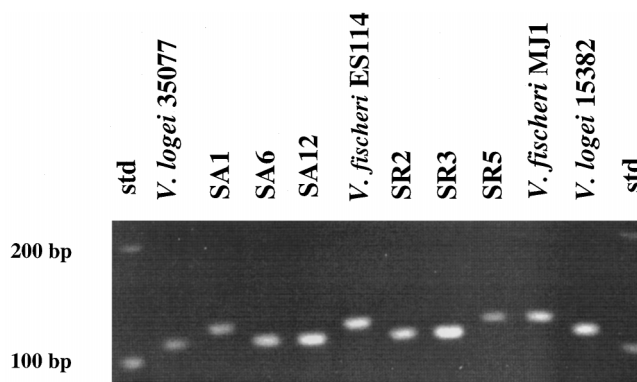


FIG. 1. Electrophoretic separation of PCR-amplified 16S rDNA fragments containing the V1 (variable) region. The names of the strains used are indicated above the lanes. std, molecular size standards.

than at 20°C; conversely, the generation times of the *V. fischeri* strains tested (SA1 and SR5) were, respectively, 1.6 and 2.1 times longer at 20°C than at 24°C.

When grown at 24 and 26°C, *V. logei* symbionts always emitted a lower level of luminescence per cell than when they were grown at 20°C (Fig. 2B). This effect was due to two temperature-sensitive processes, one that could be relieved by the addition of extraneous aliphatic aldehyde to the cells and another that could not. For instance, exposure of *V. logei* cells grown at 26°C to decyl aldehyde enhanced their luminescence between 6- and 11-fold (Table 3); however, no enhancement by decyl aldehyde was observed with *V. fischeri* symbionts from *Sepiolo* spp. Thus, the *V. logei* symbionts, as well as the ATCC strains (data not shown), appear to have a temperature-sensitive capacity for the synthesis of the natural aldehyde substrate for the luminescence reaction. In addition, even in the presence of excess aldehyde, the maximum luminescence per cell fell as much as 100-fold with increasing growth temperature between 20 and 26°C (Fig. 2B). Once again, this effect was not observed with the *V. fischeri* strains, whose luminescence per cell generally increased with increasing growth temperature from 20 to 24°C. Thus, both the growth and the luminescence of *V. fischeri* symbionts appear to be better suited to temperatures at or above 24°C, while those of *V. logei* symbionts are optimal below 24°C.

Colonization of the *E. scolopes* light organ by *Sepiolo* symbionts. Newly hatched juveniles of *E. scolopes* became luminous within 24 h of exposure to the native symbiont *V. fischeri* ES114 at water temperatures of either 21 or 26°C (Table 4). symbiotic *V. fischeri* strains from *Sepiolo* spp. also infected the

TABLE 2. Biochemical activities used to distinguish among luminous species

Strain	Biochemical activity ^a		
	Lysine decarboxylation	Nitrate reduction	D-Galactose fermentation
<i>V. logei</i> ATCC 35077	+	+	+
<i>V. fischeri</i> ES114	+	+	+
<i>V. salmonicida</i> NCMB 2262	—	—	—
SA7	+	+	+
SA12	+	+	+
SR3	+	+	+
SR6	+	+	+

^a +, positive; —, negative.

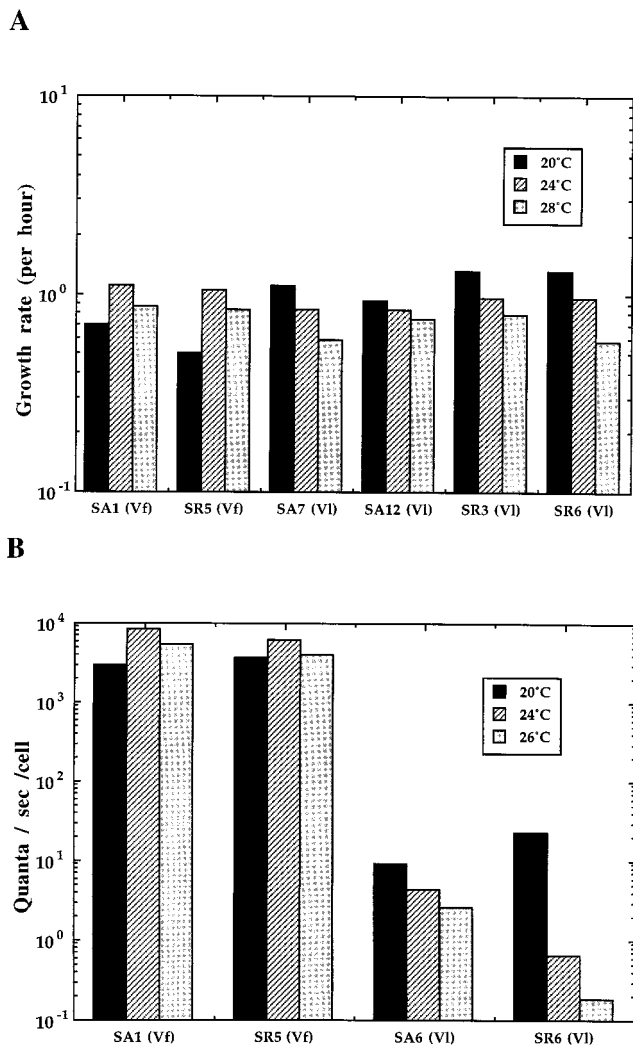


FIG. 2. (A) Growth rates (per hour) of Mediterranean *Sepiola* symbionts incubated at three temperatures. (B) Maximum luminescence per cell of exponentially growing *Sepiola* symbionts exposed to decyl aldehyde.

E. scolopes light organ with 100% efficiency at 26°C; however, the average CFU per light organ from squid infected with these strains was lower than that for squid infected with ES114. Interestingly, three of these five *V. fischeri* strains showed reduced efficiencies of colonization at 21°C (i.e., an average of only 21% of the juveniles were colonized) relative to those at 26°C.

In contrast, *V. logei* strains from *Sepiola* light organs were even less effective at colonizing juvenile *E. scolopes*. CFU were not detected in 93% of the light organ homogenates of animals exposed to strain SA5, SA6, SA7, SA12, SR3, and in those six animals that were colonized, it was to a much lower level than that of ES114. Unlike the other *V. logei* isolates, strain SR6 colonized the *E. scolopes* light organ with a high efficiency at 21°C; however, both the efficiency and extent of colonization were significantly lower at 26°C. Thus, overall, the *V. fischeri* symbionts of *Sepiola* spp. exhibited a relatively increased symbiotic competence at the higher temperature while, in contrast, the *V. logei* strains showed a relatively decreased symbiotic competence for colonization at the higher temperature.

TABLE 3. Effect of temperature and addition of decyl aldehyde on luminescence of representative *Sepiola* symbionts

Strain	Avg maximum luminescence at temp (°C) of ^a :		
	20	24	26
<i>V. fischeri</i>			
SA1	1.0	1.0	1.0
SR5	1.0	1.0	1.0
<i>V. logei</i>			
SA6	3.8	7.4	6.6
SR6	1.3	6.0	11.3

^a Average maximum luminescence of log-phase cells grown at each of three temperatures. The values given are the ratios of the specific luminescence (quanta per second per cell) of cells exposed to decyl aldehyde divided by those of cells not exposed to decyl aldehyde.

DISCUSSION

The marine luminous bacteria comprise at least 10 described species in three genera (23). Only three of these luminous species have been previously recognized as participating in specific light organ symbioses with marine animals. We report that a fourth species of luminous bacteria, *V. logei*, is the predominant light organ symbiont of *S. affinis* and *S. robusta*. Surprisingly, in some of these light organs, *V. fischeri* co-occurs as a less-abundant symbiont. Therefore, in contrast to other known bioluminescent symbioses, the light organs of *Sepiola* spp. cannot be described as symbiont species specific. While there are relatively few characteristics that can be used to separate strains of *V. fischeri* and *V. logei*, the species are nevertheless robust (2), and the symbionts isolated from the light organs of *Sepiola* species unambiguously group with one or the other of these two species. We must conclude that these symbiotic organs can be inhabited by strains of either *V. logei* or both species.

Preliminary microscopic and metabolic analyses of light organ symbionts from *S. affinis* and *S. robusta* (designated SA and SR strains, respectively) led to the conclusion that these bacteria were strains of either *V. fischeri* or *V. logei*. A similar

TABLE 4. Colonization of *E. scolopes* juveniles by symbionts from *Sepiola* spp.

Strain	CFU/squid (10^5) after 48 h at temp (°C) of ^a :	
	21	26
<i>V. fischeri</i>		
ES114	2.8 (5/5)	4.4 (11/11)
SA1	ND ^b (0/6)	1.8 (5/5)
SA2	2.4 (6/6)	2.2 (5/5)
SA3	0.2 (1/6)	0.1 (6/6)
SA4	0.7 (3/7)	0.6 (6/6)
SR5	4.4 (7/7)	3.1 (7/7)
<i>V. logei</i>		
SA5	0.9 (2/7)	ND (0/16)
SA6	ND (0/5)	ND (0/6)
SA7	0.5 (1/11)	0.3 (1/7)
SA12	ND (0/11)	ND (0/10)
SR3	0.1 (2/10)	ND (0/7)
SR6	1.0 (10/10)	0.1 (3/10)

^a Infection assays were performed at either 21 or 26°C. The numbers in parentheses are the number of animals (over the total number tested) that became luminous upon exposure to the bacterial strain.

^b ND, none detected (<14 CFU/light organ).

uncertainty exists concerning the light organ symbionts of *S. atlantica*, which have been reported to be *Photobacterium (Vibrio) fischeri*, although no information indicating how these strains were distinguished from *V. logei* was presented (13). Unfortunately, beyond the relative psychrophily of *V. logei* (Fig. 2A) (2), there are few reported phenotypic differences between these two species, reflecting their close phylogenetic relatedness (15). Nevertheless, Reichelt et al. (27), using total DNA-DNA hybridization, concluded that these two species shared only 37 to 41% DNA sequence identity, prompting them to support the assignment of separate-species status. Recently, it has been reported that a simpler means of differentiating these two species is by the presence or absence of a 10-bp gap in the V1 region of the 16S rDNA sequence (33). PCR amplification of the V1 region of the symbionts from *Sepiolo* spp. indicated that greater than 80% of them were, in fact, *V. logei* while the remaining strains were *V. fischeri* (Fig. 1). A mixed colonization population may also exist in the light organs of *S. atlantica* and perhaps other *Sepiolo* spp. (12). During the course of these analyses, we made the incidental but intriguing discovery that several luminous bacterial isolates from Southern California seawater were also *V. logei*. Thus, the population of bioluminescent bacteria found in these waters includes this species as well as the previously reported *V. fischeri* and *V. harveyi* (32).

Sequences of the 16S rRNA genes from *Sepiolo* symbiont strains SA5, SA6, SA12, and SR6 exhibited the highest degree of identity (>93%) with a *V. logei* sequence, rather than that of *V. fischeri* (87%), further supporting the results of the V1 region analyses. Interestingly, the 16S rDNA sequences from these strains also aligned to that of *V. salmonicida*, a fish pathogen that causes cold-water vibriosis in Norwegian rainbow trout and salmon, with a high degree of identity ($\geq 90\%$) (8). In addition, the *V. salmonicida* 16S rRNA gene had the same distinctive 10-bp sequence gap that was found in the *V. logei* symbionts. Despite these similarities, the presence of three metabolic activities (lysine decarboxylation, nitrate reduction, and D-galactose fermentation) in all of the SA and SR symbiont strains eliminated the possibility that these bacteria were *V. salmonicida* (Table 2). A similar ambiguity resulting from the high degree of sequence identity between *V. logei* and *V. salmonicida* may have arisen in the work of Wiik et al. (35). They described a bacterial strain designated HI651 which, despite deviating metabolically and physiologically from the *V. salmonicida* type strain NCMB 2262, was designated *V. salmonicida* on the basis of a partial shared 16S rDNA sequence identity of 99.7%. Fox et al. (11) reported that closely related *Bacillus* strains could exhibit >99.5% sequence homology yet still belong to distinct species as determined by total DNA-DNA hybridization. Thus, because the 16S rRNA gene evolves slowly relative to the rest of the genome, a considerable degree of metabolic and ecological divergence can occur between two bacterial species whose 16S rDNA sequences do not differ significantly (11). We suggest that the metabolic and physiological traits reported for strain HI651 (35) are actually more consistent with those of *V. logei*. Because *V. logei* was not included in the phylogenetic analyses of Wiik et al. (35), this hypothesis remains to be tested.

Our taxonomic studies of *Sepiolo* spp. symbionts also raise fundamental questions about the microecology and dynamics of their mixed-species light organ populations, as well as the evolution of light organ symbioses in general. Specifically, we have addressed the following three questions: (i) is the physiology of *V. logei* better adapted to cooler ambient conditions than is that of *V. fischeri*, and, thus, (ii) is temperature an important factor in determining which species of symbiont is

predominant in *Sepiolo* spp. light organs; and (iii) is there evidence that *V. logei* and *V. fischeri* symbionts of *Sepiolo* spp. have evolved sets of colonization determinants different from those expressed by the native *V. fischeri* symbiont of *E. scolopes*?

The luminescence of ATCC strains of *V. logei*, as well as that of SA and SR *V. logei* strains, was significantly reduced at warmer temperatures (>20°C). Because luminescence in these strains could be restored by exposing them to decyl aldehyde, it is likely that either the expression of the genes responsible for intrinsic aldehyde synthesis (i.e., *luxCDE* homologs [23]) or the activity of their gene products is reduced up to 11-fold at warmer growth temperatures. Such a significant temperature dependency was previously observed in the psychrophilic marine bacterium strain WSU (20). While this strain initially was reported to be *V. fischeri*, its description more closely resembles that of the subsequently established species *V. logei*. Because strain WSU, as well as the *V. logei* symbionts of *Sepiolo*, is more dependent on a low ambient temperature for luminescence than are *V. fischeri* strains (Fig. 2B), comparisons of the components of the *lux* operons of these two closely related species may provide insight into the physiological control of both the transcription of these genes and the enzymology of bacterial luminescence.

Another indication that the two species exhibit different environmental temperature optima is reflected in the relative colonization effectiveness of symbiotic *V. logei* and *V. fischeri*. *V. logei* strains from *Sepiolo* spp. more often developed a successful symbiosis with a squid host at 21°C than at 26°C, while, in contrast, *V. fischeri* strains expressed a better capacity to colonize at the higher temperature (Table 4). Because the light organs of *Sepiolo* spp. can contain a mixture of *V. logei* and *V. fischeri* (Table 1), symbiosis-competent strains of both species must be found in the Mediterranean Sea. We are tempted to conclude that when *Sepiolo* spp. are infected by a mixture of the two *Vibrio* species, the identity of the predominant symbiont present in the light organ at any particular time may be determined in part by the ambient water temperature. This conclusion is supported by the observation that the environmental temperature of the Mediterranean *Sepiolo* spp. typically varies between 10 and 22°C both with depth and over the course of the year (5, 25). Thus, one might predict that if the host were to inhabit waters with a temperature in the cooler range, the growth of *V. logei* would be favored over that of *V. fischeri* (Fig. 2A). Future experiments, using the native species of *Sepiolo* rather than *E. scolopes* as the hosts, should serve to directly address this hypothesis.

Both *E. scolopes* and *Sepiolo* spp. transmit bacterial symbionts horizontally to each new generation of squid (24, 34), and thus the newly hatched squid requires some mechanism(s) for distinguishing the correct symbiont strains from among the myriad other bacteria in seawater. Strains of *V. fischeri* isolated from the light organs of all *Euprymna* spp. tested can infect juvenile *E. scolopes*, suggesting that the symbiosis is not host species specific (24). *Sepiolo* symbionts identified as *V. fischeri* were also able to colonize the light organ of juvenile *E. scolopes*, albeit in some cases to a lesser extent than the native symbiont of the squid (Table 4). The fact that *V. logei* symbionts from *Sepiolo* spp. were capable of colonizing *E. scolopes* was unexpected because previous attempts either with *V. logei* strains ATCC 35077 and 15382 or with *V. logei* isolates from Southern California seawater had been unsuccessful (16, 21). These findings suggest that *V. logei* strains that have not been isolated from a light organ are missing one or more symbiotic determinants necessary for establishing this association. In addition, the inability of SA and SR *V. logei* strains to colonize the

E. scolopes light organ with as high a success rate as symbiotic *V. fischeri* strains, particularly at 26°C, suggests that their symbiotic determinants (29) differ sufficiently from those present in the native *E. scolopes* symbiont to account for the differences in colonization competence (Table 4).

In summary, this study reports five new conclusions. (i) For the first time, *V. logei* has been demonstrated to be a light organ symbiont, namely of the Mediterranean bobtail squids *S. robusta* and *S. affinis*. (ii) The light organs of *Sepiolo* spp. can contain a mixed colonization population consisting of a dominant species (*V. logei*) and a minor species (*V. fischeri*). (iii) The light organ symbioses of Mediterranean (*Sepiolo*) and Pacific (*Euprymna*) squid species, while arising in a common sepiolid ancestor, have evolved independently to associate with different species of *Vibrio*, perhaps influenced by the environmental temperature of the host. (iv) Luminescence in *V. logei* is temperature dependent, due in part to an aldehyde substrate limitation. (v) *V. logei* and *V. fischeri* symbionts from *Sepiolo* spp. generally exhibit diminished degrees of colonization competence compared to the native *V. fischeri* symbiont from *E. scolopes*.

ACKNOWLEDGMENTS

We thank Kyu-Ho Lee for bringing to our attention the PCR approach for differentiating *Vibrio* species and Michele Nishiguchi and Margaret McFall-Ngai for sharing unpublished data.

This research was supported by grants to E.G.R. from the Office of Naval Research (N00014-93-1-0846) and the National Institutes of Health (RR12294) and to M. McFall-Ngai and E.G.R. from the National Science Foundation (IBN96-01155).

REFERENCES

- Altschul, S. F., W. Gish, W. Miller, E. W. Meyers, and D. J. Lipman. 1990. Basic local alignment search tool. *J. Mol. Biol.* **215**:403–410.
- Bang, S. S., P. Baumann, and K. H. Nealson. 1978. Phenotypic characterization of *Photobacterium logei* (sp. nov.), a species related to *P. fischeri*. *Curr. Microbiol.* **1**:285–288.
- Baumann, P., L. Baumann, S. S. Bang, and M. J. Woolkalis. 1980. Reevaluation of the taxonomy of *Vibrio*, *Beneckeia*, and *Photobacterium*: abolition of the genus *Beneckeia*. *Curr. Microbiol.* **4**:127–132.
- Bello, G. 1995. A key for the identification of the Mediterranean sepiolids (Mollusca: Cephalopoda). *Bull. Inst. Oceanogr. Monaco* **16**:41–55.
- Bhaud, M., G. Jacques, and C. Razouls. 1967. Données météorologiques et hydrologiques de la région de Banyuls-sur-Mer, 1965–1966 (point côtier). *Vie Milieu* **18**:137–151.
- Boettcher, K. J., and E. G. Ruby. 1990. Depressed light emission by symbiotic *Vibrio fischeri* of the sepiolid squid *Euprymna scolopes*. *J. Bacteriol.* **172**:3701–3706.
- Colwell, R. R. 1965. Proposal of a neotype, ATCC 15381, for *Vibrio marinus* (Russell) Ford 1927 and request for an opinion. *Int. Bull. Bacteriol. Nomencl. Taxon.* **15**:165–175.
- Difco Laboratories Inc. 1984. *Difco manual*, 10th ed. Difco Laboratories, Inc., Detroit, Mich.
- Egidius, E., R. Wiik, K. Andersen, K. A. Hoff, and B. Hjeltnes. 1986. *Vibrio salmonicida* sp. nov., a new fish pathogen. *Int. J. Syst. Bacteriol.* **36**:518–520.
- Farmer, J. J., and F. W. Hickman-Brenner. 1991. The genera *Vibrio* and *Photobacterium*, p. 2952–3011. In A. Balows, H. G. Truper, M. Dworkin, W. Harder, and K. H. Schleifer (ed.), *The prokaryotes*, 2nd ed. Springer-Verlag, New York, N.Y.
- Fitzgerald, J. M. 1977. Classification of luminous bacteria from the light organ of the Australian pinecone fish, *Cleidopus gloriamaris*. *Arch. Microbiol.* **112**:153–156.
- Fox, G. E., J. D. Wisotzkey, and P. Jurtschuk, Jr. 1992. How close is close: 16S rRNA sequence identity may not be sufficient to guarantee species identity. *Int. J. Syst. Bacteriol.* **42**:166–170.
- Herfurth, A. H. 1936. Beiträge zur kenntnis der bakteriensymbiose der Cephalopoden. *Z. Morphol. Oekol. Tiere* **31**:561–607.
- Herring, P. J., M. R. Clarke, S. von Boletzky, and K. P. Ryan. 1981. The light organs of *Sepiolo atlantica* and *Spirula spirula* (Mollusca: Cephalopoda): bacterial and intrinsic systems in the order Sepioidea. *J. Mar. Biol. Assoc. U. K.* **61**:901–916.
- Herring, P. J., and J. G. Morin. 1978. Bioluminescence in fishes, p. 272–293. In P. J. Herring and J. G. Morin (ed.), *Bioluminescence in action*. Academic Press, New York, N.Y.
- Kita-Tsakamoto, K., H. Oyaizu, K. Nanba, and U. Simidu. 1993. Phylogenetic relationships of marine bacteria, mainly members of the family *Vibrionaceae*, determined on the basis of 16S rRNA sequences. *Int. J. Syst. Bacteriol.* **43**:8–19.
- Lee, K.-H., and E. G. Ruby. Unpublished data.
- Lee, K.-H., and E. G. Ruby. 1994. Effect of the squid host on the abundance and distribution of symbiotic *Vibrio fischeri* in nature. *Appl. Environ. Microbiol.* **60**:1565–1571.
- Leisman, G., D. Cohn, and K. H. Nealson. 1980. Bacterial origin of luminescence in marine animals. *Science* **208**:1271–1273.
- Lu, C. C., A. Guerra, F. Palumbo, and W. C. Summers. 1992. Order Sepioidea Naef, 1916. *Smithson. Contrib. Zool.* **513**:21–36.
- Makemson, J. C. 1973. Control of *in vivo* luminescence in psychrophilic marine photobacterium. *Arch. Mikrobiol.* **93**:347–358.
- McFall-Ngai, M., and E. G. Ruby. 1991. Symbiont recognition and subsequent morphogenesis as early events in an animal-bacterial mutualism. *Science* **254**:1491–1493.
- McFall-Ngai, M. J. 1990. Cypsis in the pelagic environment. *Am. Zool.* **30**:175–188.
- Nealson, K. H., and J. W. Hastings. 1991. The luminous bacteria, p. 625–639. In A. Balows, H. G. Truper, M. Dworkin, W. Harder, and K. H. Schleifer (ed.), *The prokaryotes*, 2nd ed., Springer-Verlag, New York, N.Y.
- Nishiguchi, M., and M. J. McFall-Ngai. Unpublished data.
- Panhouse, M., G. Jacques, and J. Neveux. 1975. Hydrologie dans la zone néritique de Banyuls-sur-Mer (Golfe du Lion), 1973. *Vie Milieu* **25**:77–84.
- Reichert, J. L., and P. Baumann. 1973. Taxonomy of the marine, luminous bacteria. *Arch. Mikrobiol.* **94**:283–330.
- Reichert, J. L., P. Baumann, and L. Baumann. 1976. Study of genetic relationships among marine species of the genera *Beneckeia* and *Photobacterium* by means of *in vitro* DNA/DNA hybridization. *Arch. Microbiol.* **110**:101–120.
- Reichert, J. L., K. H. Nealson, and J. W. Hastings. 1977. The specificity of symbiosis: pony fish and luminescent bacteria. *Arch. Microbiol.* **112**:157–161.
- Ruby, E. G. 1996. Lessons from a cooperative bacterial-animal association: the *Vibrio fischeri*-*Euprymna scolopes* light organ symbiosis. *Annu. Rev. Microbiol.* **50**:591–624.
- Ruby, E. G., and L. M. Asato. 1993. Growth and flagellation of *Vibrio fischeri* during initiation of the sepiolid squid light organ symbiosis. *Arch. Microbiol.* **159**:160–167.
- Ruby, E. G., and K. H. Nealson. 1976. Symbiotic associations of *Photobacterium fischeri* with the marine luminous fish *Monocentris japonica*: a model of symbiosis based on bacterial studies. *Biol. Bull.* **151**:574–586.
- Ruby, E. G., and K. H. Nealson. 1978. Seasonal changes in the species composition of luminous bacteria in nearshore seawater. *Limnol. Oceanogr.* **23**:530–533.
- Soares, C. A. G., R. C. Prado, C. B. Barros, and A. Coelho. 1995. PCR size analysis of 16S rRNA V1 region: a tool for *Vibrio* taxonomic studies, abstr. R-9, p. 480. In *Abstracts of the 95th General Meeting of the American Society for Microbiology 1995*. American Society for Microbiology, Washington, D.C.
- Wei, S. L., and R. E. Young. 1989. Development of symbiotic bacterial luminescence in a nearshore cephalopod, *Euprymna scolopes*. *Mar. Biol.* **103**:541–546.
- Wiik, R., E. Stackebrandt, O. Valle, F. L. Daae, O. M. Rødseth, and K. Andersen. 1995. Classification of fish-pathogenic vibrios based on comparative 16S rRNA analysis. *Int. J. Syst. Bacteriol.* **45**:421–428.
- Zablen, L. B. 1976. Ph.D. thesis. University of Illinois, Urbana.